

プロトプラスト融合 (Protoplast fusion) 法による二倍体細胞の取得

- 1 harvest $5 \times 10^7 - 10^8$ cells
- 2 wash with 1ml SP1
- 3 incubate in SP1 for 5 min at room temperature.
- 4 spin down, and suspend with 500 μ l SP2 (containing 10 mg/ml Novozyme (Sigma L1412-5G))
- 5 incubate for 30 min at 37°C

*Avoid too long incubation in this step not to lyse the cells.

*After the treatment of Novozyme here, cells should be pelleted by 3000 rpm for 3 min, or by チビタン for several seconds.

- 6 mix 2 μ l cell suspension with equal volume of water on a slide glass and check protoplasts under a microscope (protoplast will burst by adding water).

*If protoplasts are not observed, add newly prepared SP2 containing Novozyme, and incubate for 30 min.

- 7 pellet and wash cells with 1ml SP3 for 2 times

*Suspend cells moderately by pipeting.

*Although lysed cells are sometimes difficult to be pelleted, replacing SP2 with SP3 make it easy to pellet the lysed cells.

- 8 suspend cells in SP4, adjust cell concentration to $5 \times 10^7/50 \mu$ l
- 9 mix 50 μ l of each strains into one tube, suspend moderately.
- 10 add 1ml SP5, incubate for more than 30 min at 26-30°C
- 11 pellet and suspend cells in 100-200 μ l SP4.
- 12 spread cells on selective medium containing 1.2 M sorbitol.

Solutions

SP1 (SP2+EDTA)

1.2M D-sorbitol

50mM Citrate-phosphate (pH 5.6)

40mM EDTA

SP2

1.2M D-sorbitol

50mM Citrate-phosphate (pH 5.6)

SP3

1.2M D-sorbitol

10mM Tris-HCl (pH 7.6)

SP4 (SP3+CaCl₂)

1.2M D-sorbitol

10mM Tris-HCl (pH 7.6)

10mM CaCl₂

SP5

20% PEG4000

10mM Tris-HCl (pH 7.6)

10mM CaCl₂